

# The sound of many hands clapping

Tumultuous applause can transform itself into waves of synchronized clapping.

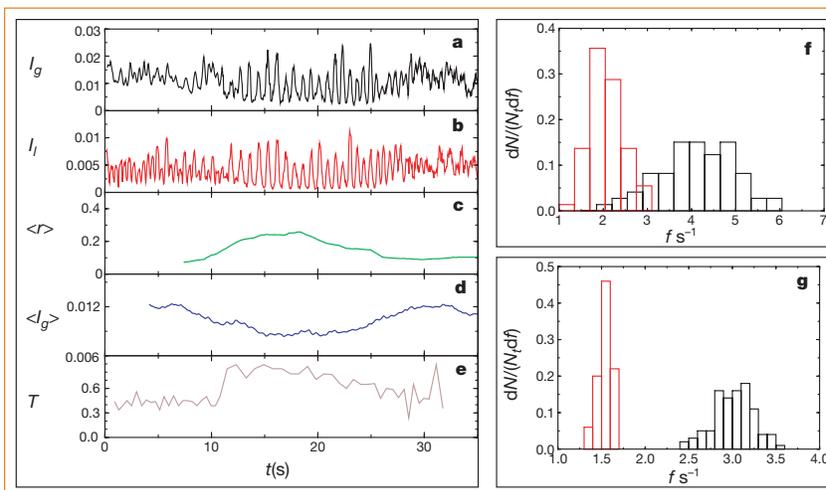
An audience expresses appreciation for a good performance by the strength and nature of its applause. The thunder of applause at the start often turns quite suddenly into synchronized clapping, and this synchronization can disappear and reappear several times during the applause. The phenomenon is a delightful expression of social self-organization that provides an example on a human scale of the synchronization processes that occur in numerous natural systems, ranging from flashing Asian fireflies to oscillating chemical reactions<sup>1–3</sup>. Here we explain the dynamics of this rhythmic applause.

We investigated the mechanism and development of synchronized clapping by making a series of measurements focusing on the collective aspects of the self-organization process as well as on the behaviour of the individuals in the audience. We recorded several theatre and opera performances in Romania and Hungary by using a microphone placed on the ceiling of the hall (Fig. 1a). Typically, after a few seconds of incoherent random clapping, a periodic signal developed (a sign of synchronized clapping), visible in Fig. 1a as pronounced spikes. The transition is also captured by the order parameter (Fig. 1c), which increases as the periodic signal develops, and decreases as it disappears.

Although synchronization increases the strength of the signal at the moment of the clapping, it leads to a decrease in the average noise intensity in the room (Fig. 1d). This is surprising, as the driving force for synchronization would be expected to reflect the desire of the audience to express its enthusiasm by increasing the average noise intensity.

The origin of this conflict between the average noise and synchronization can be understood by correlating the global signal with the behaviour of an individual in the audience. We demonstrated this by recording the local sound intensity in the vicinity of a group of oblivious individuals (Fig. 1b). In the incoherent phase, the local signal was periodic, with a short period corresponding to the fast clapping of an individual in the audience. However, the clapping period suddenly doubled at the start of the synchronized phase (at about 12 seconds in Fig. 1a,b), and slowly decreased as synchronization was lost (Fig. 1e).

The decrease in the average noise intensity is therefore a consequence of the period doubling, because there is less clapping per unit time. An increase in the average noise intensity is possible only by decreasing the



**Figure 1** Emergence of synchronization in clapping. **a**, Global noise intensity ( $I_g$ ) as a function of time. The digitized data were squared and the moving average was determined over a window of size 0.2 s, several times shorter than the clapping period. A characteristic region indicates the appearance and disappearance of the synchronized clapping. Over several performances, we recorded 50 similar sequences of synchronized clapping (for additional data sets and audio recordings, see <http://www.nd.edu/~networks/clap>). **b**, Local noise intensity ( $I_l$ ), measured by a hidden microphone in the vicinity of a spectator. **c**, Order parameter,  $r$ , defined as the maximum of the normalized correlation between the signal  $c(t)$  and a harmonic function,  $r = [\max_{\phi, \omega} \int_{t-T}^{t+T} c(t) \sin(2\pi/T + \phi) dt] / [\int_{t-T}^{t+T} c(t) dt]$ , where  $\phi$  and  $T$  span all possible values. **d**, Average noise intensity, obtained by taking a moving average over a 3-s window of the global noise intensity shown in **a**. **e**, The clapping period,  $T$ , defined as the intervals between the clearly distinguishable maxima. **f**, The normalized histogram of clapping frequencies measured for 73 high-school students (isolated from each other) for mode I (black) and mode II (red) clapping. **g**, Normalized histogram for mode I and II clapping obtained for a single student, sampled 100 times over a one-week period.

clapping period, which can indeed occur (Fig. 1e). However, the decreasing clapping period gradually brings the synchronized clapping back to the fast clapping heard in the early asynchronous phase, and synchronization disappears. Apparently, this conflicting desire of the audience simultaneously to increase the average noise intensity and to maintain synchronization leads to the sequence of appearing and disappearing synchronized regimes.

These results indicate that the transition from random to synchronized clapping is accompanied by a period-doubling process. To determine whether period doubling is in fact a necessary condition for synchronization, we investigated the internal frequency of clapping by several individuals in controlled clapping experiments. Individual students, isolated in a room, were instructed to clap as they would immediately after a good performance (mode I clapping) or during the rhythmic applause (mode II clapping).

The frequencies of the two modes of clapping are clearly separated and the average period doubles from mode I to mode II clapping (Fig. 1f). Most important, however, is that the width of the frequency distribution and the relative dispersion of mode II clapping is considerably smaller, a result

that is reproducible for a single individual as well (Fig. 1g).

Our results indicate that after an initial asynchronous phase, characterized by high-frequency clapping (mode I), individuals synchronize by eliminating every second beat, suddenly shifting to a clapping mode with a double period (mode II) where dispersion is smaller. For a group of globally coupled oscillators, the condition for synchronization is that dispersion must be smaller than a critical value<sup>4,5</sup>. Consequently, period doubling emerges as a condition for synchronization, because it leads to slower clapping modes during which significantly smaller dispersion can be maintained.

Our measurements offer an insight into the mechanism of synchronized clapping: during fast clapping, synchronization is not possible owing to the large dispersion in the clapping frequencies. After period doubling, as mode II clapping with small dispersion appears, synchronization can be and is achieved. However, as the audience gradually decreases the period to enhance the average noise intensity, it slips back to the fast clapping mode with larger dispersion, destroying synchronization.

In summary, the individuals in the audience have to be aware that by doubling their clapping period they can achieve synchro-

nization. This perhaps explains why in the smaller and culturally more homogeneous eastern European communities, synchronized clapping is a daily event, whereas it happens only sporadically in western European and North American audiences. In general, our results offer evidence of a novel route to synchronization, not yet observed in physical or biological systems<sup>2,3,5,6</sup>.

Z. Néda\*, E. Ravasz\*, Y. Brechet†, T. Vicsek‡, A.-L. Barabási§

\* Babeş-Bolyai University, Department of Theoretical Physics, Str. Kogălniceanu 1, RO-3400 Cluj-Napoca, Romania

† ENSEEG-LTPCM, INP-Grenoble, Saint-Martin D'Herès, Cedex, France

‡ Department of Biological Physics, Eötvös-Loránd University, Budapest 1117, Hungary

§ Department of Physics, University of Notre Dame, Notre Dame, Indiana 46556, USA  
e-mail: zned@phys.ubbcluj.ro

1. Strogatz, S. H. & Stewart, I. *Sci. Am.* 102–109 (December, 1993).
2. Bottani, S. *Phys. Rev. E* 54, 2334–2350 (1997).
3. Glass, L. & Mackey, M. C. *From Clocks to Chaos: The Rhythms of Life* (Princeton Univ. Press, Princeton, NJ, 1988).
4. Winfree, A. T. *J. Theor. Biol.* 16, 15 (1967).
5. Kuramoto, Y. & Nishikawa, I. *J. Stat. Phys.* 49, 569–605 (1987).
6. Mirollo, R. & Strogatz, S. H. *SIAM J. Appl. Math.* 50, 1645–1662 (1990).

Hormones

Leptin and diabetes in lipoatrophic mice

Lipoatrophic (lipodystrophic) diabetes is a disorder in which insulin resistance and hyperglycaemia are associated with a reduced body-fat mass<sup>1</sup>, in contrast to the usual association of diabetes with obesity. Transgenic mice with differing degrees of fat loss can be used as models for lipoatrophy<sup>2–4</sup>. Using the aP2-SREBP-1c mouse<sup>3</sup>, which has a moderate fat deficiency, Shimomura *et al.* showed that leptin treatment reverses the diabetes, concluding that insulin resistance in congenital generalized lipodystrophy can be explained by a leptin deficiency<sup>5</sup>. However, we have used a more severe model of lipoatrophy, the A-ZIP/F-1 mouse<sup>2,6</sup>, in which we find that leptin treatment is only slightly effective in correcting diabetes.

A-ZIP/F-1 mice (Table 1) have an almost complete lack of white adipose (fat) tissue, a severe resistance to insulin, diabetes, and greatly reduced serum leptin levels<sup>2</sup>. We found that infusing leptin into A-ZIP/F-1 mice at the same rate (5 µg day<sup>-1</sup> for 4 weeks, starting at 7 weeks of age) and to produce the same serum leptin level (3 ng ml<sup>-1</sup>) as in Shimomura *et al.*'s mice had no effect on serum glucose or insulin concentrations (results not shown). A higher leptin dose (30 µg per day, causing leptin to rise by 5 ng ml<sup>-1</sup>) did reduce glucose and insulin levels (Fig. 1), food intake (from 6.6 ± 0.3 to 4.7 ± 0.2 g day<sup>-1</sup>, P < 0.001), and liver weight (from 3.04 ± 0.04 to 2.09 ± 0.11 g, P < 0.001). Even at this higher dose, however, our mice still had markedly raised blood glucose and insulin levels.

In our A-ZIP/F-1 mice, the efficacy of

Figure 1 Effect of continuous subcutaneous leptin infusion on serum glucose and insulin concentrations. Leptin (R&D Systems) was infused into male mice by using an osmotic pump (Alzet) at 30 µg day<sup>-1</sup> for six days, ending at 7 or 13 weeks of age, as indicated. Data are mean and s.e.m. with n = 6/group; single and double asterisks indicate differences between treated and control at P ≤ 0.01 and P < 0.001, respectively. Non-fasting glucose was measured using a Glucometer Elite (Bayer) and insulin was quantified by radioimmunoassay (Linco).

leptin treatment diminished with age: at 13 weeks (the age of Shimomura *et al.*'s mice), leptin had a minimal effect (Fig. 1), and no effect at all at 28 weeks (results not shown). In contrast, leptin infusion into 13-week-old leptin-deficient ob/ob mice completely normalized both glucose and insulin levels (Fig. 1).

Evidence from humans and mice supports the conclusion that leptin deficiency cannot completely explain the diabetic phenotype of generalized lipoatrophy. Patients with generalized lipoatrophy are more

prone to diabetes<sup>1</sup> than are those who lack leptin<sup>7–9</sup>; similarly, A-ZIP/F-1 mice are more diabetic than ob/ob mice (Fig. 1). Thus leptin deficiency contributes to the insulin resistance of generalized lipoatrophy, but is neither the sole nor the principal cause of insulin resistance in severe forms of this disease.

The observed differences between the A-ZIP/F-1 and aP2-SREBP-1c mice are probably due to their different amounts of fat, although transgene-specific effects or their different genetic backgrounds may play a part. aP2-SREBP-1c mice have more residual adipose tissue: in these mice, leptin appears to be limiting and its replacement reverses their diabetes. A-ZIP/F-1 mice must experience loss of other functions provided by adipose tissue besides leptin secretion — for example, functions that affect fatty-acid and triglyceride metabolism. Alternatively, adipose tissue might exert a direct or indirect endocrine effect.

Oksana Gavrilova\*, Bernice Marcus-Samuels\*, Lisa R. Leon\*, Charles Vinson†, Marc L. Reitman\*

\* Diabetes Branch, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Bethesda, Maryland 20892, USA

e-mail: mlr@helix.nih.gov

† Laboratory of Metabolism, National Cancer Institute, National Institutes of Health, Bethesda, Maryland 20892, USA

1. Foster, D. W. in *Harrison's Principles of Internal Medicine* (eds Isselbacher, K. J. *et al.*) 2131–2136 (McGraw-Hill, New York, 1994).
2. Moitra, J. *et al. Genes Dev.* 12, 3168–3181 (1998).
3. Shimomura, I. *et al. Genes Dev.* 12, 3182–3194 (1998).
4. Burant, C. F. *et al. J. Clin. Invest.* 100, 2900–2908 (1997).
5. Shimomura, I., Hammer, R. E., Ikemoto, S., Brown, M. S. & Goldstein, J. L. *Nature* 401, 73–76 (1999).
6. Gavrilova, O. *et al. Proc. Natl Acad. Sci. USA* 96, 14623–14628 (1999).
7. Montague, C. T. *et al. Nature* 387, 903–908 (1997).
8. Strobel, A., Issad, T., Camoin, L., Ozata, M. & Strosberg, A. D. *Nature Genet.* 18, 213–215 (1998).
9. Farooqi, I. S. *et al. N. Engl. J. Med.* 341, 879–884 (1999).

Shimomura *et al. reply* — Human lipodystrophy (also called lipoatrophic diabetes) is genetically heterogeneous, with the severity of insulin resistance and diabetes mellitus varying widely depending on the degree of reduction in adipose tissue mass and the age of the patient<sup>1,2</sup>. It is therefore not surprising that two mouse models of lipodystrophy (created by using two different transgenes, A-ZIP/F-1 and aP2-SREBP-1c) vary in their disease severity and in their sensitivity to leptin. The aP2-SREBP-1c animals respond to leptin with a decrease in their insulin and blood sugar levels<sup>3</sup>, whereas the A-ZIP/F-1 animals of Gavrilova *et al.* apparently manifest leptin resistance. The differences between these two models should not preclude a clinical trial of leptin in leptin-deficient patients with lipodystrophy, with continuation of therapy in those who are leptin-sensitive.

Table 1 Differences in the severity of the A-ZIP/F-1 and aP2-SREBP-1c phenotypes

	Epididymal fat mass (% of wild type)	Brown fat mass (% of wild type)	Glucose (mg dl <sup>-1</sup> , non-fasting)	Insulin (ng ml <sup>-1</sup> , non-fasting)
A-ZIP/F-1	≤1	~50	~1,000	~60
aP2-SREBP-1c, line A (ref. 3)	~30	~400	~300	~20

Data are for 7-week-old mice.